



I. B. Anweting<sup>1</sup>, S. O. Idris<sup>2</sup> and A. D. Onu<sup>3</sup>

<sup>1</sup>Department of Chemistry, University of Uyo, Akwa Ibom State, Nigeria

<sup>2</sup>Department of Chemistry, Ahmadu Bello University, Zaria-Nigeria

<sup>3</sup>Department of Chemistry, Federal College of Education, Zaria-Nigeria

\*Corresponding author: [law4chem@gmail.com](mailto:law4chem@gmail.com)

Received: June 27, 2020 Accepted: November 13, 2020

**Abstract:** Redox reaction of tetrakis(2,2'- bipyridine)- $\mu$ -oxodiiron(III) complex,  $\text{Fe}_2\text{O}^{4+}$  and glutathione (GSH) has been carried out in aqueous hydrochloric acid. The reaction was carried out at  $[\text{H}^+] = 0.001 \text{ mol dm}^{-3}$ ,  $I = 0.3 \text{ mol dm}^{-3}$  (NaCl),  $T = 27 \pm 1^\circ\text{C}$ , and  $\lambda_{\text{max}} = 520 \text{ nm}$ . One mole of oxidant is consumed per mole of reductant. The reaction is first order with respect to  $[\text{Fe}_2\text{O}^{4+}]$  and zero order on  $[\text{GSH}]$  and is not hydrogen ion and ionic strength dependent. Added anions and cations have no effect on the reaction; moreover there was no gel formation when acrylamide and excess methanol were added to the reaction mixture, which shows the absence of polymerization. With recourse to experimental data, the reaction is rationalised to follow outer sphere mechanism with ion pair character.

**Keywords:** Kinetics, mechanism, redox, glutathione, oxidant and reductant

### Introduction

Glutathione (GSH) also known as  $\gamma$ -L-glutamyl-L-cysteinylglycine is a thiol with -SH functional group at cysteine. GSH is required for several cell processes interconnected with alterations in the maintenance and regulation of the thiol-redox status, due to its capability to exist in different redox specie (Forman *et al.*, 2009). GSH is a major antioxidant in the brain (Dringen, 2000), with a concentration which is much higher than that in blood or cerebrospinal fluid (Cooper and Kristal, 1997). It exerts its functions via several mechanisms. First, GSH non-enzymatically reacts with superoxide (Winterbourn and Metodiewa, 1994), NO (Clancy, 1994), hydroxyl radical (Bains and Shaw, 1997), and ONOO<sup>-</sup> (Koppal *et al.*, 1999).

In particular, GSH has a higher ability to scavenge superoxide than cysteine (Hussaini *et al.*, 1996). Furthermore, there is no known enzymatic defense against hydroxyl radicals, making GSH the only compound capable of scavenging these radicals (Bains and Shaw, 1997). GSH also serves as an essential cofactor for a number of enzymes. It works as an electron donor for the reduction of  $\text{H}_2\text{O}_2$  or other peroxides catalysed by glutathione peroxidases (Chance *et al.*, 1979). The brain has a relatively high level of glutathione peroxidases as compared with that of catalase, while the liver has high levels of both (Maher, 2005). GSH reacts with various endogenous and xenobiotic compounds mediated by glutathione-S-transferase (GST) (Commandeuret *et al.*, 1995) to form mixed disulphides, which are exported to the outside of the cell. GSH can also react with 4-hydroxynonenal via the action of GST to form the GSH-hydroxynonenal adduct (Xie *et al.*, 1998). This process plays an important role in cellular detoxification. Moreover, GSH is the major redox buffer and maintains intracellular redoxhomeostasis. Under conditions of oxidative stress, GSH can lead to the reversible formation of mixed disulphides between protein thiol groups (S-glutathionylation), a process critical for preventing irreversible oxidation of proteins (Giustarini *et al.*, 2004). GSH is thought to exert dual (agonistic/antagonistic) actions on neuronal responses mediated by NMDA receptors in the brain. GSH also serves as an endogenous NO reservoir to form S-nitrosoglutathione (Singh *et al.*, 1996).

The chemistry of diiron complexes for the past decades have continued to be of great interest because of the presence of such diiron centres in a variety of non-haem iron proteins. Participation of the  $\mu$ -oxo diiron cores of the metalloproteins, hemerythrin, ribonucleotide reductase, and purple acid

phosphatase in their biological oxygen-transport and oxygenation processes is well known (Stenkamp *et al.*, 1981; Anatanaitis and Aisen, 1983; Sjöberg and Graslund 1983; Wilkins and Harrington, 1983; Sheriff *et al.*, 1987). Though the diiron complexes have been known for long, to a large extent, greater focus was on their synthesis and characterisation. Numerous diiron(III) complexes have been prepared and characterised using several spectroscopic techniques such as infrared, UV-visible and nuclear magnetic resonance (Gaines *et al.*, 1936; Schugar *et al.*, 1967, 1969; Reiff *et al.*, 1968; David *et al.*, 1972; Reiff, 1977; Nozaki *et al.*, 1999).

In view of the roles GSH, as enumerated above, there is need for kinetic data of the electron transfer of this important biochemical with tetrakis (2,2'- bipyridine)- $\mu$ - oxodiiron(III) complex. The kinetic data generated from the electron transfer reaction between the biochemical compound and  $\text{Fe}_2\text{O}^{4+}$  will complement the much needed kinetic information and will bring to the limelight their electron transfer properties.

### Materials and Methods

#### Experimentals

##### Materials

Tetrakis (2,2'- bipyridine)- $\mu$ -oxo-diiron(III) chloride ( $[\text{Fe}_2(\text{bpy})_4\text{O}]\text{Cl}_4$ ) hereafter referred to as  $\text{Fe}_2\text{O}^{4+}$  was prepared, purified and characterized following the method of (David, 1973). A stock solution of GSH (Sigma-Aldrich) was prepared by dissolving appropriate quantity of GSH into distilled water in volumetric flask and made up to the mark. Stock solutions of  $2.0 \text{ mol dm}^{-3}$  (HCl) was made by diluting 8.5 ml of 36% HCl (specific gravity 1.18) in 50 ml standard flask, then made up to the mark with distilled water. The solution was standardised titrimetrically with standard solution of previously dried  $\text{Na}_2\text{CO}_3$  using methyl red as indicator (Chimere *et al.*, 1985). Stock solutions of sodium chloride, sodium sulphate, sodium acetate, potassium chloride and magnesium chloride were prepared from analar grade salts and their various concentrations were obtained by serial dilution.

##### Stoichiometry

The stoichiometry of the reactions was determined by spectrophotometric titration at  $\lambda_{\text{max}} = 520 \text{ nm}$  using the mole ratio method (Ukoha and Iyun, 2001, 2002). The concentration of  $\text{Fe}_2\text{O}^{4+}$  was kept constant at  $6.7 \times 10^{-5} \text{ mol dm}^{-3}$  while that of GSH was varied between (2.68 – 29.5)  $\times 10^{-5} \text{ mol dm}^{-3}$  at  $[\text{H}^+] = 1.0 \times 10^{-3} \text{ mol dm}^{-3}$  and constant ionic

strength of  $0.3 \text{ mol dm}^{-3}$  (NaCl) at  $T = 27 \pm 1.0^\circ\text{C}$ . The reactions were allowed to stand until the repeated absorbances of the reaction mixture at  $\lambda_{\text{max}} = 520 \text{ nm}$  were constant. The stoichiometry was then determined from the plot of absorbance versus mole ratio of  $\text{Fe}_2\text{O}^{4+}$ : GSH.

**Kinetic studies**

The rate of reaction was studied under pseudo-first order condition with [GSH] in at least 40 folds excess over  $[\text{Fe}_2\text{O}^{4+}]$  at the stated conditions by monitoring the increase in the absorbance of the complex at  $520 \text{ nm}$  using Corning Colorimeter 525. From the slopes of pseudo-first order plots of  $\log(A_\infty - A_t)$  versus time, the pseudo-first order rate constant ( $k_1$ ) were determined.

**Acid dependence studies**

The effect of changes in the hydrogen ion concentration on the reaction rate was investigated by keeping the concentration of the other reactants constant while varying the hydrogen ion concentration in the range  $(4 - 14) \times 10^{-4} \text{ mol dm}^{-3}$ .

**Effect of ionic strength**

The effect of ionic strength on the rates of the reaction was studied over a range of  $(1.0 - 6.0) \times 10^{-1} \text{ mol dm}^{-3}$  using NaCl, while others reaction conditions were kept constant.

**Influence of added anions**

The influence of added sulphate and acetate ions on the rate of reaction were investigated by varying the concentration of these anions while keeping  $[\text{Fe}_2\text{O}^{4+}]$ , [GSH] and ionic strength constant.

**Test for participation of free radicals in the course of reaction**

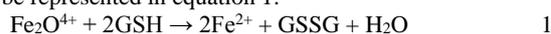
Test for free radicals was carried out by adding 2 g of acrylamide to a partially oxidised reaction mixture containing various concentrations of oxidant, reductant and hydrogen ion. A large excess of methanol was added to the reaction mixture. Control experiment was carried out by adding acrylamide to solutions of oxidant and reductant separately at the same conditions of  $[\text{H}^+]$ , I and temperature. Any polymerisation as indicated by gel formation suggested the presence of free radicals in the reaction mixture (Iyun and Adegite, 1990; Vaidya *et al.*, 1991).

**Products analysis**

At the completion of the reaction, the reaction mixtures were analysed for the type of organic and inorganic products formed. The test for the presence of disulphide was carried out according to literature (McAuley and Gomwalk, 1968, 1969). The GSH reacted with a little excess of the oxidant in acid medium and ionic strength of reaction. At the completion of the reaction, the mixture was extracted six times with diethyl ether. The combined ether extracts were washed and dried with anhydrous  $\text{Na}_2\text{SO}_4$  and left overnight to dry.

**RESULTS AND DISCUSSION**

Stoichiometric studies showed that one mole of  $\text{Fe}_2\text{O}^{4+}$  is reduced per two moles of GSH oxidised. The overall reactions can be represented in equation 1.



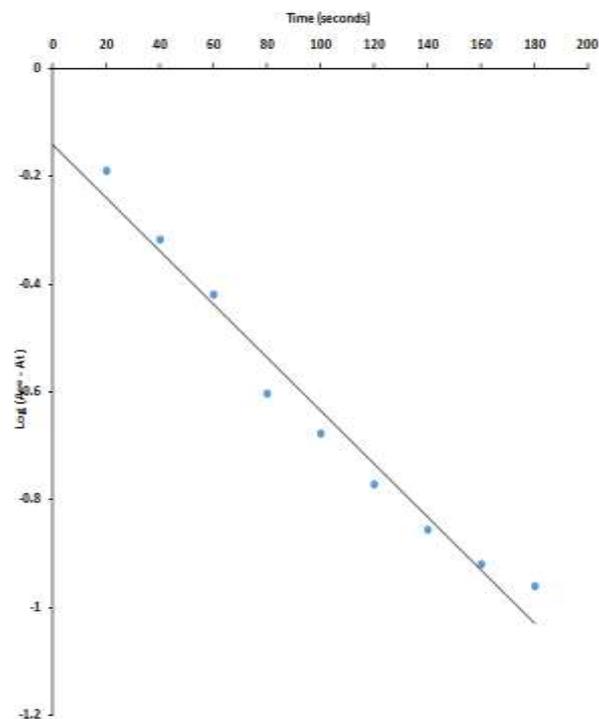
The stoichiometry of 1:2 (oxidant: reductant) is in agreement with the report given by other authors as follows:  $\text{Ru}_2\text{O}^{4+}$  with glutathione (Ayoko *et al.*, 1993), Iron(III)-2,2'-bipyridyl with methionine (Tiruvedhula *et al.*, 1995),  $\text{Ru}_2\text{O}^{4+}$  with L-cysteine (Iyun *et al.*, 1996),  $[(\text{FeHEDTA})_2\text{O}]^{2-}$  with mercaptoacetic acid, mercaptoethylamine and mercaptoethanol (Ukoha, 1999; Ukoha and Iyun, 2001). However, this mole ratio is not analogous to 1:1 (oxidant: reductant) reported for oxidation of mercaptoacetic acid by 12-tungstocobaltate(III) (Ayoko, 1981; Ayoko and Olatunji, 1983), tris (polypyridyl) iron(III) complex with glutathione (Ayoko *et al.*, 1993), reaction of trisoxalato cobaltate(III) iron with mercaptoacetic acid

(Lawal *et al.*, 1994),  $\text{Ru}_2\text{O}^{4+}$  with mercaptoethylamine and mercaptoethanol (Iyun *et al.*, 1995c),  $\text{Ru}_2\text{O}^{4+}$  with mercaptoacetic acid (Musa *et al.*, 1998), oxidation of mercaptoacetic acid and L-cysteine by  $\text{Fe}_2\text{O}^{4+}$  (Idris *et al.*, 2004; Idris, 2005).

Tiruvedhula *et al.* (1995) reported that the oxidation of many thiols can give rise to many products depending on the nature of the oxidant. Methionine and mercaptobenzoic acid were oxidised to sulphoxide by oxidants such as chloroauric acid, Cr(IV), hexachloroiridate sulphite, iron (III) -2, 2'-bipyridyl and  $\text{Mn}^{\text{II}}\text{O}_2\text{M}^{\text{IV}}$  (Natile *et al.*, 1976; Goswami *et al.*, 1981; Olatunji and Ayoko, 1988; Ayoko *et al.*, 1992; Tiruvedhula *et al.*, 1995; Lohdip, 1999). In the reaction of  $\text{Fe}_2\text{O}^{4+}$  with mercaptoacetic acid and L-cysteine, derivative of sulphonic acid was obtained as the organic product (Idris *et al.*, 2004; Idris, 2005). However disulphide (GSSG) was formed as the only organic product in the reactions and this was confirmed using the method of McAuley and Gomwalk (1968 and 1969). Subsequently the crystals obtained from the above method was subjected to FTIR spectroscopy to authenticate the formation of disulphide (GSSG). The weak absorption at  $542 \text{ cm}^{-1}$  confirmed the formation of disulphide (GSSG). Disulphide formation has been observed in the reactions of  $\text{Ru}_2\text{O}^{4+}$  with L-cysteine (Iyun *et al.*, 1996), and mercaptoacetic acid (Musa *et al.*, 1998) and  $[(\text{FeHEDTA})_2\text{O}]^{2-}$  with mercaptoacetic acid, mercaptoethanol and mercaptoethylamine (Ukoha 1999; Ukoha and Iyun, 2001). Addition of  $\text{K}_3[\text{Fe}(\text{CN})_6]$  to the products of the systems gave a blue black colour which depicts the presence of  $\text{Fe}^{2+}$  as an inorganic product.

Kinetic studies of the reduction of  $\text{Fe}_2\text{O}^{4+}$  by GSH indicated first order dependence on the  $[\text{Fe}_2\text{O}^{4+}]$ . Pseudo-first order plots of  $\log(A_\infty - A_t)$  versus time were linear (Fig. 1) for about 85% extent of reaction. The invariance of  $k_{\text{obs}}$  in Table 1 indicated zero order dependence on the [GSH]. The rate of redox process can therefore be represented as;

$$\frac{1}{2} \frac{d[\text{Fe}^{2+}]}{dt} = k_{\text{obs}}[\text{Fe}_2\text{O}^{4+}] \quad (2)$$



**Fig. 1:** Typical Pseudo-first order plot for the redox reaction of  $\text{Fe}_2\text{O}^{4+}$  with GSH at  $[\text{Fe}_2\text{O}^{4+}] = 2.0 \times 10^{-4} \text{ mol dm}^{-3}$ ,  $[\text{GSH}] = 5.0 \times 10^{-3} \text{ mol dm}^{-3}$ ,  $[\text{H}^+] = 1.0 \times 10^{-3} \text{ mol dm}^{-3}$ ,  $I = 0.30 \text{ mol dm}^{-3}$  (NaCl),  $T = 26.0 \pm 1.0^\circ\text{C}$  and  $\lambda_{\text{max}} = 520 \text{ nm}$

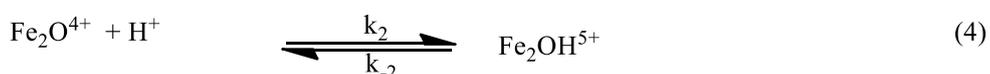
**Table 1: Pseudo-first order rate constants for the redox reaction of  $\text{Fe}_2\text{O}^{4+}$  – GSH in aqueous HCl medium,  $\lambda_{\text{max}} = 520 \text{ nm}$ ,  $I = 0.30 \text{ mol dm}^{-3}$  (NaCl),  $[\text{Fe}_2\text{O}^{4+}] = 5 \times 10^{-5} \text{ mol dm}^{-3}$ ,  $T = 26.0 \pm 1.0^\circ\text{C}$**

$10^3[\text{GSH}]$ ( $\text{mol dm}^{-3}$ )	$10^2[\text{H}^+]$ ( $\text{mol dm}^{-3}$ )	$10 [\text{I}]$ ( $\text{mol dm}^{-3}$ )	$10^3 k_{\text{obs}}$ ( $\text{s}^{-1}$ )
2.0	1.0	3.0	4.32
4.0	1.0	3.0	3.92
6.0	1.0	3.0	4.10
8.0	1.0	3.0	4.08
12.0	1.0	3.0	4.01
14.0	1.0	3.0	4.01
16.0	1.0	3.0	3.89
6.0	0.4	3.0	4.13
6.0	0.6	3.0	4.36
6.0	0.8	3.0	4.24
6.0	1.0	3.0	4.40
6.0	1.2	3.0	3.87
6.0	1.4	3.0	4.23
6.0	1.0	1.0	3.60
6.0	1.0	2.0	4.12
6.0	1.0	3.0	4.10
6.0	1.0	4.0	3.70
6.0	1.0	5.0	3.70
6.0	1.0	6.0	4.01

The zeroth order with respect to [GSH] is similar to what was obtained in the reduction of  $\text{Fe}_2\text{O}^{4+}$  by mercaptoacetic acid and L-cysteine (Idris, 2005). However, this result does not conform to the result from earlier researchers. For instance, first order dependence were observed on both [reductant] and [oxidant] in the oxidation of mercaptoethanol, mercaptoethylamine and mercaptoacetic acid by  $\text{Ru}_2\text{O}^{4+}$  (Iyun *et al.*, 1995c; Musa *et al.*, 1998) and  $[(\text{FeHEDTA})_2\text{O}]^{2-}$  (Ukoha, 1999; Ukoha and Iyun, 2001), in the reactions of Cr(VI) with L-cysteine (Iyun and Tinouye, 1998), mercaptoacetic acid by  $\text{IO}_3^-$  (Ukoha and Ibrahim, 2004) and mononuclear Fe(III) with thiols (Wiberg *et al.*, 1968). Within the acid range of  $(4.0 - 14) \times 10^{-4} \text{ mol dm}^{-3}$  the rate of reaction displayed non-dependence on  $[\text{H}^+]$  in the reaction of  $\text{Fe}_2\text{O}^{4+}$  and GSH. Similar report has been posited

in the reaction of the oxidant ( $\text{Fe}_2\text{O}^{4+}$ ) with mercaptoacetic acid and L-cysteine (Idris, 2005). On the other hand, increased in reaction rate with increase in  $[\text{H}^+]$  has been reported in the reaction of thiols with trisoxalatocobaltate(III) ion (Lawal *et al.*, 1994) and trispolypyridyl iron(III) complexes (Ekubo, 1992; Ayoko *et al.*, 1993a, b), also inverse acid dependence in thiols reaction with  $\text{Ru}_2\text{O}^{4+}$  (Ayoko *et al.*, 1993; Iyun *et al.*, 1995, 1996; Musa *et al.*, 1998), iron(III)-2,2'-bipyridyl (Tiruveedhula *et al.*, 1995), Cr(VI) (Iyun and Tinouye 1998), and  $[(\text{FeHEDTA})_2\text{O}]^{2-}$  (Ukoha, 1999; Ukoha and Iyun, 2001) have been reported. Varying the ionic strength of the reaction media between 0.1 – 0.6  $\text{mol dm}^{-3}$  (NaCl), had no effect on the rates of reaction. Change in the dielectric constant of the medium did not affect the rate of the reaction as well. Similar observation with respect to the effect of ionic strength and dielectric constant on the reaction rate have been noticed in the reaction of  $[(\text{FeHEDTA})_2\text{O}]^{2-}$  with mercaptoethylamine and mercaptoethanol (Ukoha and Iyun, 2001) and  $\text{Fe}_2\text{O}^{4+}$  with mercaptoacetic acid and L-cysteine (Idris, 2005). Nonetheless reactions of mercaptoacetic acid and L-cysteine with  $\text{Ru}_2\text{O}^{4+}$  (Iyun *et al.*, 1996; Musa *et al.*, 1998) decreased with increase in ionic strength. Also there was a marked enhancement of the rate of these reactions as a function of  $1/D$ , both of these features posited that, the rate determining step involved oppositely charged redox species.

Free radical test was carried out by addition of acrylamide to the reaction mixtures, followed by excess methanol, there was no gel formation, suggesting that polymerisation has not occurred. Lack of polymerization from this reaction suggests probable absence of free radical formation during the electron transfer. On the other hand, free radical could have been formed but reacts so quickly that this method could not detect it (Iyun *et al.*, 1995). The rate of reaction was not affected by added cations and anions. Absence of spectroscopic evidence for the formation of intermediate during the reaction suggests that a precursor complex is probably not formed prior to the act of electron transfer and that the electron transfer may occur by the outer-sphere path. With recourse to the empirical kinetic data, the mechanism of this reaction is proposed as follows:



$$\text{Rate} = k_4[\text{Fe}_2\text{OH}^{5+}, \text{GSGS}^{2-}] \quad (9)$$

Application of steady state hypothesis for  $[\text{Fe}_2\text{OH}^{5+}, \text{GSGS}^{2-}]$  gives:

$$[\text{Fe}_2\text{OH}^{5+}, \text{GSGS}^{2-}] = \frac{k_3[\text{Fe}_2\text{OH}^{5+}][\text{GSH.GS}^-]}{k_{-3}[\text{H}^+] + k_4} \quad (10)$$

If  $k_{-3}[\text{H}^+] \gg k_4$  equation 10 reduces to

$$[\text{Fe}_2\text{OH}^{5+}, \text{GSGS}^{2-}] = \frac{k_3[\text{Fe}_2\text{OH}^{5+}][\text{GSH.GS}^-]}{k_{-3}[\text{H}^+]} \quad (11)$$

Application of steady state hypothesis for  $[\text{Fe}_2\text{OH}^{5+}]$  gives

$$[\text{Fe}_2\text{OH}^{5+}] = \frac{k_2[\text{Fe}_2\text{O}^{4+}][\text{H}^+]}{k_{-2} + k_3[\text{GSH.GS}^-]} \quad (12)$$

From equation 12, if  $k_{-2} \ll k_3[\text{GSH.GS}^-]$ , then the equation reduces to:

$$[\text{Fe}_2\text{OH}^{5+}] = \frac{k_2[\text{Fe}_2\text{O}^{4+}][\text{H}^+]}{k_3[\text{GSH.GS}^-]} \quad (13)$$

Substituting equation 13 into 11 gives:

$$[\text{Fe}_2\text{OH}^{5+}, \text{GSGS}^{2-}] = \frac{k_3 k_2 [\text{Fe}_2\text{O}^{4+}][\text{H}^+][\text{GSH.GS}^-]}{k_{-3}[\text{H}^+] k_3[\text{GSH.GS}^-]} \quad (14)$$

$$[\text{Fe}_2\text{OH}^{5+}, \text{GSGS}^{2-}] = \frac{k_2}{k_{-3}} [\text{Fe}_2\text{O}^{4+}] \quad (15)$$

Substituting equation 15 into 9 gives:

$$\text{Rate} = \frac{k_4 k_2}{k_{-3}} [\text{Fe}_2\text{O}^{4+}] \quad (16)$$

$$\text{Thus } \frac{k_4 k_2}{k_{-3}} = k_{\text{obs}} \quad (17)$$

#### Conflict of Interest

Authors declare that there is no conflict of interest related to this work.

#### References

- Anatanaitis BC & Aisen P 1983. *In Binding Protein without Cofactors or Sulphur Cluster* (Theil, EC, Eichorn GL & Marzilli LG (Ed)), Elsevier, New York, 5: 111-136.
- Ayoko GA, Iyun JF & Ekubo AT 1993. Kinetics of oxidation of glutathione by tris(polypyridyl) iron(III) complexes in aqueous solution. *Indian Journal of Chemistry*, 32A: 616-618.
- Ayoko GA 1981. Reactions of 12- tungstocobaltate(III) ion with some organic and inorganic ligands in aqueous solution. M. Sc Thesis, Ahmadu Bello University, Zaria – Nigeria and The References Therein.
- Ayoko GA & Olatunji MA 1983. Oxidation of of reduction-cysteine, mercaptoacetic acid, and β-mercaptoethylamine by 12-tungstocobaltate(III). *Polyhedron*, 12: 577-582.
- Ayoko GA, Iyun JF & Mamman S 1993. Oxidation of N-(2-hydroxyethyl) ethylenediamine triacetate by tris poly(pyridyl)iron(III) complexes and the dodecatungsto cobaltate(III)iron. *Transition Metal Chemistry*, 18: 475-477.
- Ayoko GA, Iyun JF & Ekubo AT 1992. Kinetics of reduction hexachloroiridate(IV) by L-methionine in aqueous solution. *Indian Journal of Chemistry*, 31A:975-983.
- Bains JS & Shaw CA 1997. Neurodegenerative disorders in humans: The role of glutathione in oxidation stress-mediated neuronal death. *Brain Research Reviews*, 25:335-358.
- Chance B, Sies H & Boveris A 1979. Hydroperoxide metabolism in mammalian organs. *Physiological Reviews*, 59: 529-605.
- Chimere I, Mohammed A & Emmanuel J 1985. *Laboratory Exercise in Chemistry*. United industries and shipping INC, Taipei, Taiwan, pp. 79 – 85.
- Clancy RM, Levartovsky D, Leszczynska-Piziat J, Yegundin J & Abramson SB 1994. Nitric oxide reacts with intracellular glutathione and activates the hexose

monophosphate shunt in human neutrophils: evidence for S-nitrosoglutathione as a bioactive intermediary. *Proceedings of National: Academy Science USA*, 91: 3680-3684.

- Commandeur JN, Stijntjes GJ & Vermeulen NP 1995. Enzymes and transport systems involved in the formation and disposition of glutathione S-conjugates. Role in bioactivation and detoxification mechanism of xerobiotics. *Pharmacological Reviews*, 47: 271-330.
- Cooper, A.J. and Kristal, B.S. (1997). Multiple roles of glutathione and the central nervous system. *Biological Chemistry*, 378: 793-802.
- David PG & De Mello PC 1973. Kinetics of the dissociation of binuclear oxgen-bridged complexes of iron(III) with 1,10-phenanthroline and 2,2' bipyridine. *Inorganic Chemistry*, 12(9): 2188-2192.
- David PG, Richardson JG & Wehry EL 1972. Photoreduction of tetrakis (1,10-phenanthroline)-μ- oxo diiron(III) complexes in aqueous and acetonitrile solution. *J. Inorg. and Nuclear Chem.*, 34: 1333-1346.
- Dringen R 2000. Metabolism and functions of glutathione in brain. *Progress in Neurobiology*, 62: 649-671.
- Ekubo AT 1992. Kinetics and mechanisms of the oxidation of L-methionine and glutathione by some transition metals complexes. M.Sc Thesis Ahmadu Bello University, Zaria, Nigeria and the references therein.
- Forman HJ, Zhang H & Rinna A 2009. Glutathione: overview of its protective roles and measurement and biosynthesis. *Molecular Aspect of Medicine*, 30: 1-12.
- Gains A, Hammett LP & Walden GH 1936. The structure and properties of mononuclear and polynuclear phenanthroline-ferric complexes. *J. Amer. Chem. Soc.*, 58: 1688.
- Giustarini D, Rossi R, Milzani A, Colombo R & Dalle-Donne I 2004. S-glutathione. from redox regulation of protein functions to human disease. *Journal of Cellular and Moleccular Medicine*, 8: 201-212.
- Goswami, K.B., Chandr, G. and Srivastava, L. (1994). Kinetics of chronic acid oxidation of serine, methionine and cysteine. *Indian J. Chem. Soc.*, 58: 525.
- Hussaini S, Slikker W & Ali SF 1996. Role of metallothione in glutathione and other antidioxidants in scavenging superoxide radicals and their possible role in neuro protection. *Neurochemistry International*, 29: 145-152.
- Idris SO 2005. Some electron transfer reactions of Cr(VI) and of tetrakis(2,2'- bipyridine)-μ-oxodiiron(III) complex, *Ph.D. Thesis*. Chemistry Department, Ahmadu Bello University, Zaria and the references therein.
- Idris SO, Iyun JF & Agbaji EB 2004. Kinetics and mechanism of the oxidation of catechol by  $\text{Fe}_2(\text{bipy})_4\text{O}^{4+}$  in aqueous hydrochloric acid medium. *Chemclass Journal*, 29-30.
- Iyun JF, Ayoko GA & Lawal HM 1996. Kinetics of the reduction of μ-oxobis [aquobis(2, 2'- bipyridine)] ruthenium(III) by L-cysteine in aqueous solution. *Indian Journal of Chemistry*, 35(A): 210 – 213.
- Iyun JF, Musa KY & Ayoko GA 1995. Oxidation of 2 - mercaptoethanol and 2 - mercaptoethylamine by  $[(\text{bpy})_2\text{H}_2\text{O}]\text{Ru}^{\text{III}}\text{O}^{4+}$  in Aqueous Media. *Indian Journal of Chemistry*, 34(A): 635 – 638.
- Iyun JF & Tinuoye EO 1998. Chromium oxidation of L-cysteine. A proceeding of the first *Chemclass Conference Organised by the Chemical Society of Nigeria*, Zaria Chapter, pp. 56-59.
- Iyun JF & Adegite A 1990. Kinetics and mechanism of the reduction of dichlorotetrakis(2, 2-bipyridine)-μ-oxodiruthenium ion by Ti(III)-EDTA in aqueous Acidic Medium. *Bulletin of Chem. Soc. Ethiopia*, 4: 27-31.
- Koppal T, Drake J, Yatins S, Jordan B, Varadarajan S & Bettenhausen L 1999. Peroxynitrite-induced alterations

- in synaptosomal membrane, proteins: Insight into oxidative stress in Alzheimer's disease. *Journal of Neurochemistry*, 72: 310-317.
- Lawal HM, Olagbemi OT & Iyun JF 1994. Oxidation of mercaptoacetic by trisoxolato-cobaltate(III) iron in transfer reactions of *N*-methyl thiourea and diaquotetrakis(2, 2'-bipyridine)- $\mu$ -oxodiruthenium (III) ion in aqueous perchloric acid. *J. Chem. Soc. Nig.*, 19: 118-122.
- Lohdip YN 1999. Construction and evaluation of a solid state photometer and the study of the redox reactions of a mixed-valence manganese complex and of some oxyanions as models for redox active photosynthetic enzymes and biochemical system. Unpublished *Ph.D Thesis*, Ahmadu Bello University, Zaria, Nigeria.
- McAuley A & Gomwalk UD 1968. Metal ion oxidations in solution. Part V. Cerium(IV) oxidation of thiourea and its *N*-substituted derivatives. *J. Chem. Soc.*, 2948-2951.
- McAuley A & Gomwalk UD 1969. Metal ion oxidations in solution. Part VI. Oxidation of thiourea and its *N*-substituted derivatives by cobalt(III). *J. Chem. Soc. (A)*, 977-991.
- Maher P 2005. The effects of stress and ageing on glutathione metabolism. *Ageing Research Reviews*, 4: 288-314.
- Musa KY, Iyun JF & Ayoko GA 1998. Kinetics and mechanisms of reduction of diaquotetrakis(2, 2'-bipyridine)- $\mu$ -oxodiruthenium(III) ion by mercaptoacetic acid. A proceeding of the first *Chem. Conf. Organised by the Chem. Soc. Nig.*, Zaria Chapter, 51-55.
- Nozaki C, Kiyoto I, Minai Y, Misono M & Mizuno N 1999. Synthesis and characterization of diiron-substituted silicotungstate. *Inorganic Chemistry*, 38(25): 5724-5729.
- Olatunji MA & Ayoko GA 1988. Kinetics and mechanism of oxidation of L-methionine by aqueous solution of chromium(VI). *Polyhedron*, 7: 11.
- Reiff WM, Bakier (Jr), WA & Erickson NE 1968. Studies of some binuclear oxygen bridged complexes of iron (III). New iron (III) 2,2,2'-terpyridine complexes. *J. Amer. Chem. Soc.*, 90(18): 4794-4800.
- Reiff WM 1977. Strongly coupled oxo-bridged iron(III) complexes magnetically perturbed mossbauer spectra. *Journal of Chemical Physics*, 54(11): 4718-4722.
- Schugar H, Wallings C, Jones RB & Gray HB 1967. The structure of iron(III) in aqueous solution. *J. Amer. Chem. Soc.*, 89: 3712-3714.
- Schugar HJ, Hubbard AT, Anson FG & Gray HB 1969. Electrochemical and spectral studies of dimeric iron (III) complexes. *J. Amer. Chem. Soc.*, 91(1): 71-73.
- Sheriff S, Hendrickson WA & Smith JL 1987. Structure of myohemerythrin in the azidomet state at 1.7/1.3. A resolution. *Journal of Molecular Biology*, 197(2): 273-296.
- Singh RJ, Hogg N, Joseph J & Kalyanaraman B 1996. Mechanism of nitric oxide release from S-nitrosothiols. *Journal of Biological Chemistry*, 271: 18596-18603.
- Sjoberg BM & Grashund A 1983. In *Advances in Inorganic Biochemistry*, (Theil, E. C., Eichorn, G. L. and Marzilli, L.G., (Ed), Elsevier, New York, 5: 87-110
- Stenkamp RE, Sieker LC, Jensen LH & Sanders-Leohr J 1981. Structure of binuclear iron complex in metazido haemerythrin from the mistle dyscritum at 2.2.  $A^{\circ}$  resolution. *Nature*, 291: 263-264.
- Tiruveedhula RR, Parvataneni V & Lanka 1995. Mechanism of oxidation of DL-methionine by iron(III)-2,2'-bipyridyl in perchloric acid. A kinetic approach. *Transition Metal*, 20: 170.
- Ukoha PO 1999. Kinetics and mechanisms of some redox reactions of  $\mu$ -oxo-bridged iron(III) complex ion  $[(FeHEDTA)_2O]^{2-}$  and some oxyanions and thiols. Unpublished *Ph.D Thesis*, Ahmadu Bello University, Zaria, Nigeria and the references therein.
- Ukoha PO & Iyun JF 2001. Kinetics of reduction of an Iron(III) complex ion by mercaptoethanol and mercaptoethylamine in perchloric acid medium. *J. Chem. Soc. Nig.*, 26(2): 163-168.
- Ukoha PO & Iyun JF 2002. Oxidation of L-ascorbic acid by  $enH_2[(FeHEDTA)_2O].6H_2O$  in aqueous medium. *J. Chem. Soc. Nig.*, 27(2): 119-122.
- Ukoha PO & Ibrahim E 2004. Mechanism of the oxidation of  $\beta$ -mercaptoacetic acid by trioxiodate(V) in aqueous acid medium. *Chemclass Journal*, 138-141.
- Vaidya VK, Pitlia RL, Kabra BV, Mali SL & Ameta SC 1991. Dye-sensitized photo-oxidation of thiourea by singlet oxygen. *J. Photochem. and Photobiology A*, 60(1): 47-50.
- Wiberg, K. B., Maltz, H. and Ukaro, M. (1968). Mechanisms of ferricyanide oxidation of thiols. *Inorganic Chemistry*, 7:830.
- Wilkins RG & Harrington PC 1983. The Chemistry of hemerythrin. *Advances in Organic Biochemistry*, 5: 51-85.
- Winterbourn CC & Metodiewa D 1994. The reaction of superoxide with reduced glutathione. *Archive of Biochemical Biophysics*, 314: 284-290.
- Xie C, Lovell MA & Markesbery WR 1998. Glutathione transferase protects neuronal cultures against four hydroxyphenol toxicity. *Free Radical Biology and Medicine*, 25: 979-988.